



# **Bio-based technologies for microalgae harvesting**

**Saba Khalatbari, Ville-Hermann Sotaniemi, Tiina Leiviskä**

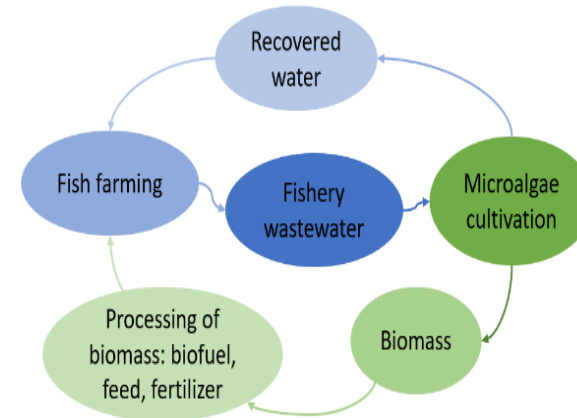
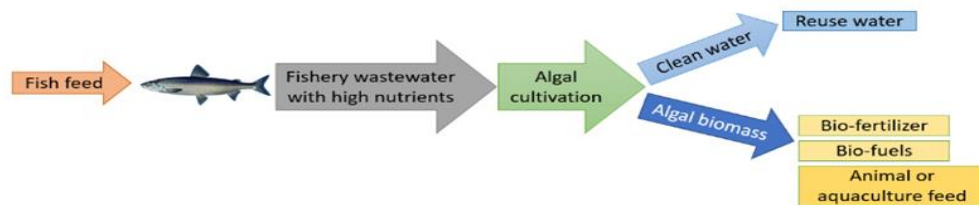
**Chemical Process Engineering Research Unit, University of Oulu**

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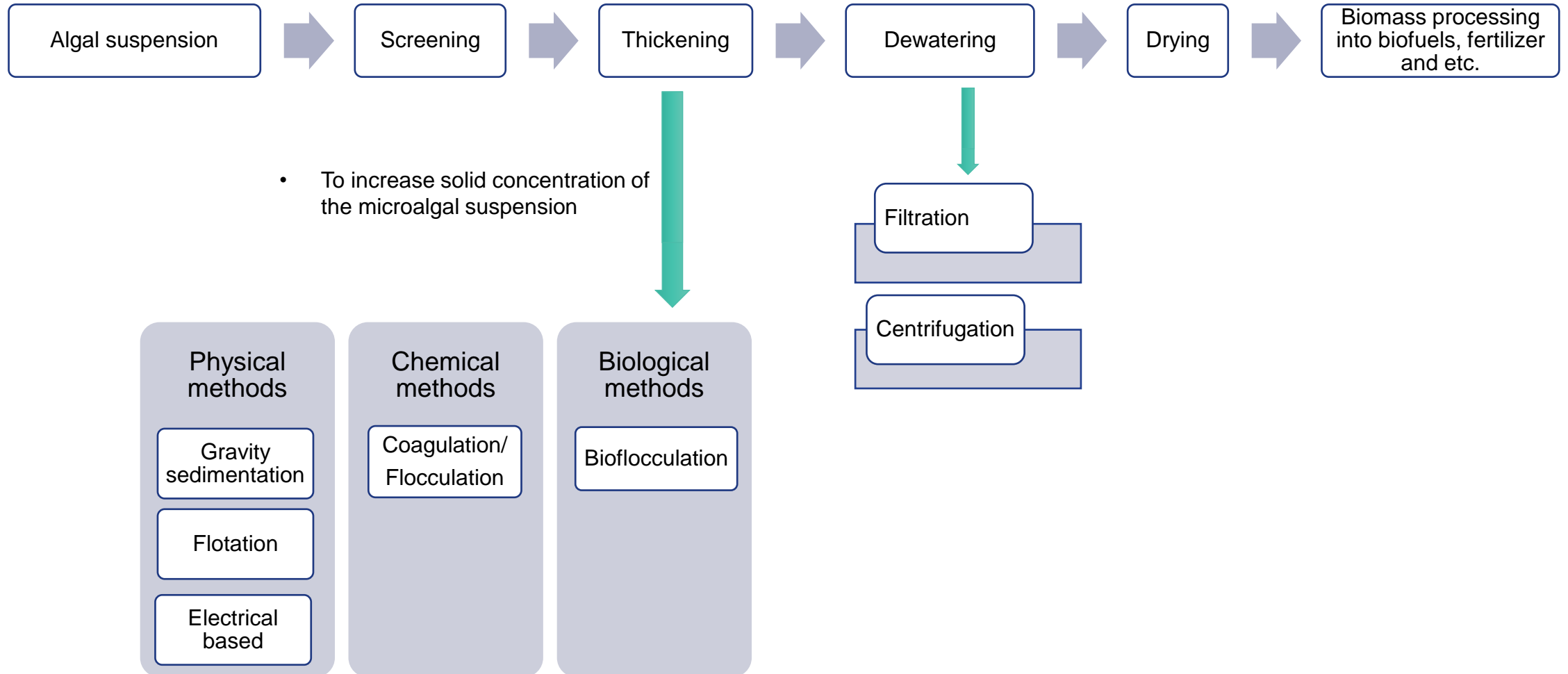
## Integration microalgae production with wastewater treatment (WWT)

- Microalgae can remove phosphate and nitrogen from wastewater.
- These nutrients can be absorbed by algal cell biomass and then treat wastewater.
- Algae cultivation in wastewater has the dual benefit of cleaning the wastewater while also producing algal biomass.
- Therefore, efficient biomass recovery or harvesting of diverse algal species represents a critical bottleneck for large-scale algal biorefinery process.





# Microalgae harvesting methods:

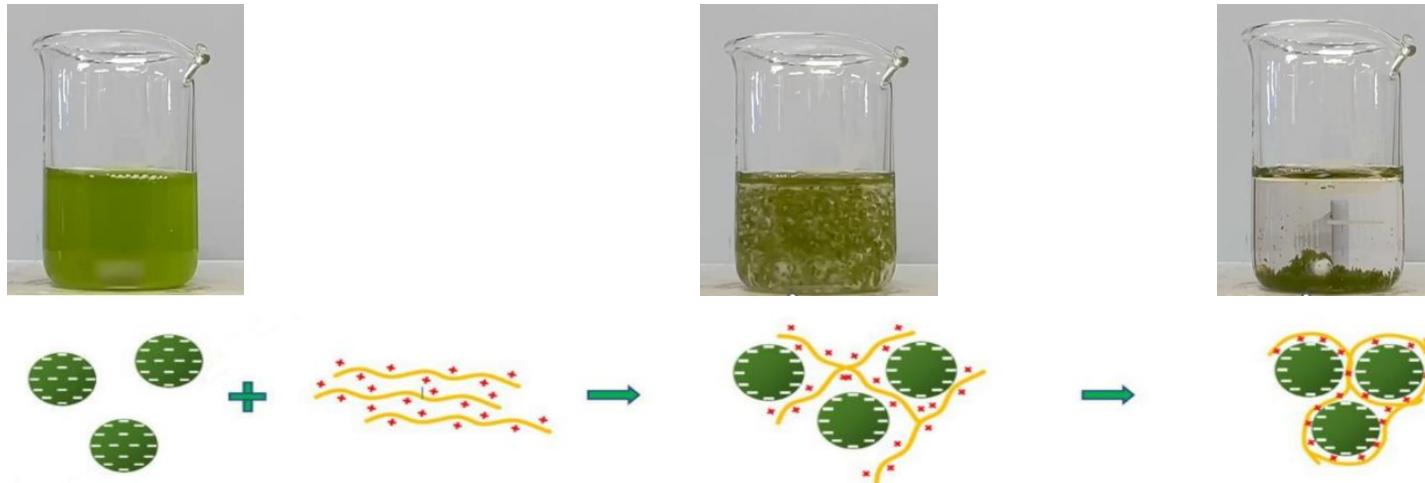


Harvesting method	Description	Advantages	Disadvantages
Gravity sedimentation	Microalgae settle by to gravity.	<ul style="list-style-type: none"> <li>Simple and inexpensive.</li> </ul>	<ul style="list-style-type: none"> <li>Time-consuming.</li> <li>Possibility of biomass deterioration.</li> <li>Low concentration of the algal cake.</li> </ul>
Flotation	Gas bubbles fed to the broth provide the lifting force needed for particle transport and separation	<ul style="list-style-type: none"> <li>Feasible for large scale applications.</li> <li>Low-cost method.</li> <li>Short operation times.</li> </ul>	<ul style="list-style-type: none"> <li>Requires the use of chemical flocculants.</li> <li>Unfeasible for marine microalgae harvesting.</li> </ul>
Electrical based processes	An electrical field is applied to the culture broth	<ul style="list-style-type: none"> <li>Applicable to a wide variety of microalgal species.</li> <li>Do not require the addition of chemical flocculants</li> </ul>	<ul style="list-style-type: none"> <li>Poorly disseminated.</li> <li>High energy and equipment costs.</li> </ul>
Chemical coagulation/flocculation	Use of chemical reagents (organic (chitosan, tannin) or inorganic (Alum, iron-based) to destabilise and increase the size of the particles	<ul style="list-style-type: none"> <li>Simple and fast method.</li> <li>Low energy requirements.</li> </ul>	<ul style="list-style-type: none"> <li>Can be expensive.</li> <li>Recycling of culture medium is limited.</li> </ul>
Auto and bioflocculation	<ul style="list-style-type: none"> <li>Flocculation by pH increase</li> <li>Flocculation by fungi, bacteria, yeast, algae, extracellular polymer</li> </ul>	<ul style="list-style-type: none"> <li>Inexpensive method.</li> <li>Allows culture medium recycling.</li> <li>Non-toxic to microalgal biomass.</li> </ul>	<ul style="list-style-type: none"> <li>Changes in cellular composition.</li> <li>Possibility of microbiological contamination.</li> </ul>
Filtration	Microalgal deposits on the filtration membrane tend to grow thicker throughout the process	<ul style="list-style-type: none"> <li>High recovery efficiencies.</li> <li>Allows the separation of shear sensitive species.</li> </ul>	<ul style="list-style-type: none"> <li>The possibility of fouling/clogging</li> <li>Regularly cleaned/replaced</li> </ul>
Centrifugation	Spinning algal broth at high speed	<ul style="list-style-type: none"> <li>Fast method.</li> <li>High recovery efficiencies.</li> </ul>	<ul style="list-style-type: none"> <li>Expensive method.</li> <li>High energy requirements.</li> <li>Possibility of cell damage due to high shear forces.</li> </ul>



## Coagulation-flocculation harvesting process of algal cells :

- Coagulation is charges' neutralization and flocculation refers to agglomeration of unstable and small particles through surface charge neutralization, electrostatic patching and/or bridging.
- Floccs formation allows for separation (or recovery) by simple gravity-induced settling or any other conventional separation method.
- The process is simple and efficient and has been extensively investigated as a promising strategy for harvesting various algal species.
- Optical Density Measurement: how much light is absorbed or scattered by a solution. In this context, optical density can provide information about the concentration of particles or flocs present in the treated suspension.





## Jar Test Setup:

### Rapid Mixing:

- Agitation speed set at around 200 rpm for 30 seconds.
- Purpose: Ensures uniform distribution of coagulant throughout the suspension.

### Coagulant Addition:

- Coagulant was added during rapid mixing phase.

### Slow Mixing:

- Agitation speed reduced to approximately 20 rpm for 10 minutes.
- Objective: Encourages the formation of larger flocs from the coagulated particles.

### Settling Phase:

- 30-minute period with no agitation.
- Force of gravity causes larger flocs to settle towards the bottom.
- Extract samples from the liquid's surface for measuring optical density.



The video playback speed was increased to 2x.

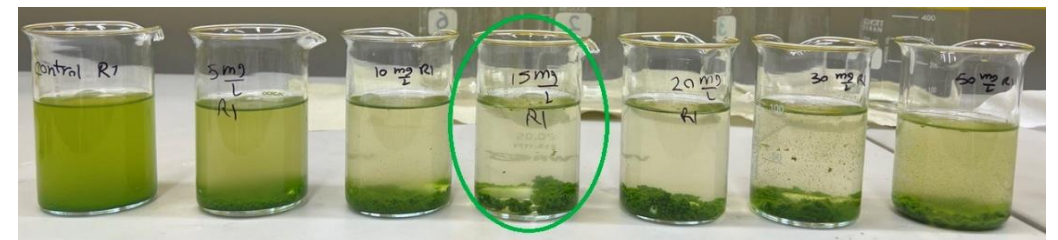
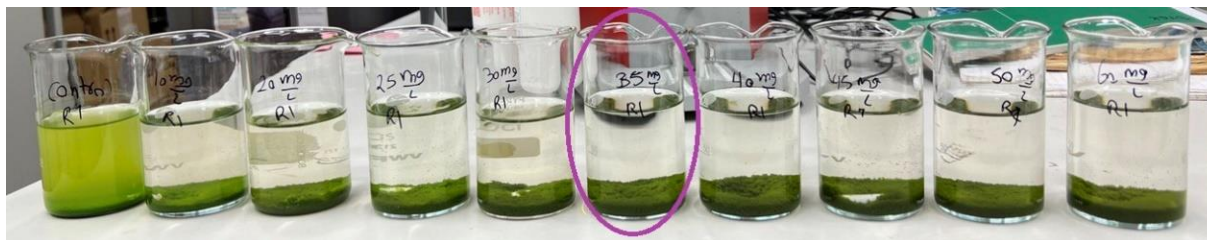


## Chemical flocculation with tannin and chitosan - optimum dosage:

### Sample Collection:

- Microalgae samples (Chlorella.S) obtained from Micropolis Oy for testing.
- A 100 ml suspension of microalgae placed in a beaker as the testing medium.

**Result: Optimum dosage of chitosan was 15 mg/l and for tannin was 35 mg/l with harvesting efficiency of 97% and 99%, respectively.**







## Chemical flocculation with tannin and chitosan - settling rate:

### Sample Collection:

- Chlorella.S microalgae samples sourced from Micropolis Oy.
- 800 ml of microalgae suspension poured into a 1000 ml beaker.

**Result: At optimal doses, settling rates were similar with both coagulants.**







## Effect of pH on microalgae flocculation

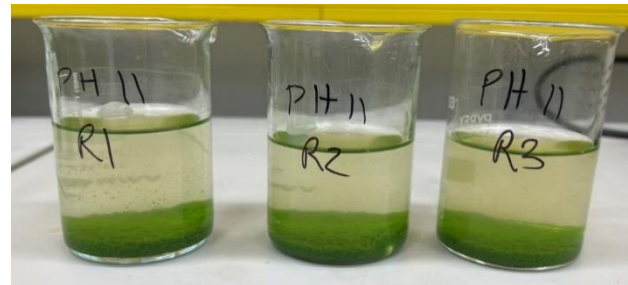
### Sample Collection:

- Chlorella.S microalgae samples sourced from Micropolis Oy.

### pH Adjustment:

- Used 1M HCl (hydrochloric acid) and 1M NaOH (sodium hydroxide) solutions.
- pH adjusted to the desired level for experimental conditions.

**Result: Higher pH increased auto-flocculation of algae (but can affect the dissolution of protein – pH adjustment not feasible)**





## **Chemical flocculation with tannin after fish processing wastewater treatment with microalgae:**

### **Microalgae was added to fish processing wastewater:**

- Microalgae initially added to fish processing wastewater.
- Optical density measurements taken after 10 and 15 days of microalgae introduction.

### **Evaluation timeline:**

#### **Day 10 and Day 15 evaluation:**

- Wastewater samples analyzed on day 10 and day 15 after tannin coagulation.
- Comparison of characteristics before and after coagulant addition.

**Results: Optimum tannin dosage was 60 mg/l with optical density of 0.066 and 0.058 on day 10 and day 15, respectively.**



## Bio-flocculation with fungus:

### Mutual Enhancement:

- Fungi consumes carbon produced by the microalgae in the medium via photosynthesis.
- Microalgae are provided with protection by the fungi.

### Fungal Strain (*F. solani*):

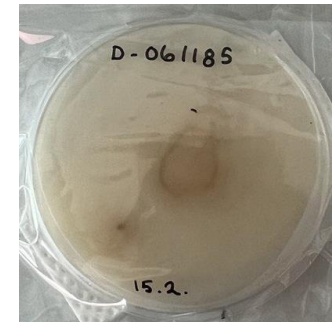
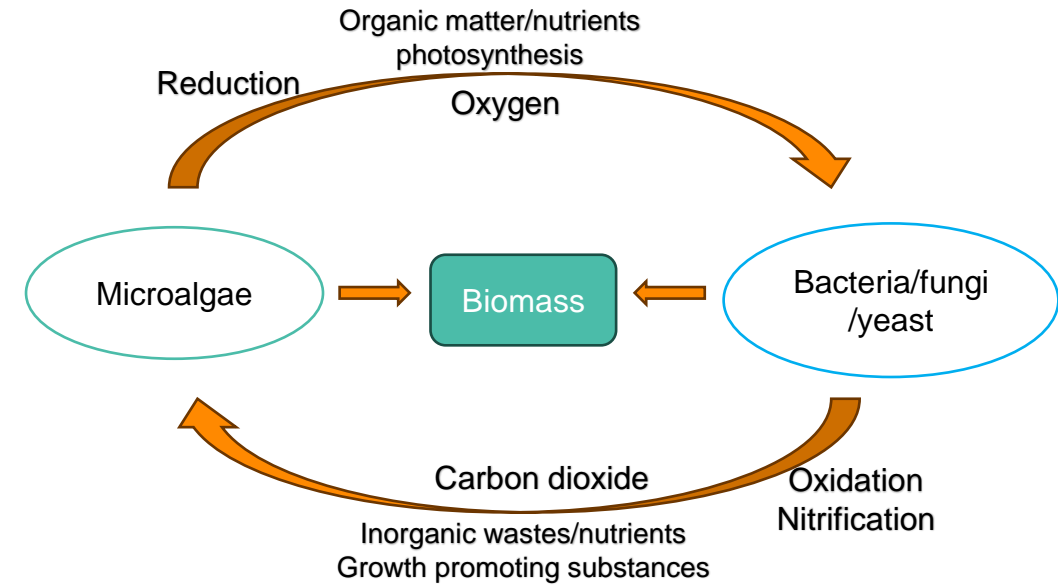
- Sourced from VTT culture collection, Finland.
- Cultivated for stability on plates and in bottles.

### Experiment Focus:

- Studied microalgae/fungi and wastewater/fungi symbiosis.
- Investigated potential benefits for wastewater treatment.

### Future Pathways:

- Building on co-culture insights.
- Exploring wider applications and process refinement.





## Synergic effect between fungus and microalgae – adding carbon source:

### Enhancing Collaboration:

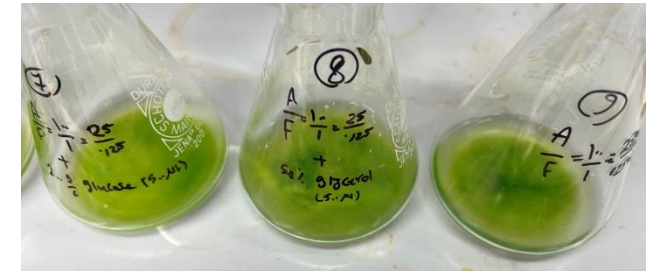
- Investigating the synergy between fungi and microalgae with the ratio of 1/100.
- Introducing additional carbon source (glucose and glycerol) to optimize effects.

### Mutual Benefits:

- Microalgae use added carbon for growth and photosynthesis.
- Fungi benefit from enhanced carbon availability.

**Result: Adding glycerol as a carbon source could enhance the synergic between algae and fungus.**

Name	TN	TP
<b>Control (Chlorella.S)</b>	<b>18.8</b>	<b>33.3</b>
A/F= 100/1	21.5	34
A/F= 100/1 + glucose	8.34	31.3
<b>A/F= 100/1 + glycerol</b>	<b>6.36</b>	<b>29.6</b>



## Synergic effect between fungus and wastewater – adding carbon source:

### Investigating Synergy:

- Exploring combined impact of fungus and wastewater.
- Focusing on the addition of a carbon source.

### Beneficial Symbiosis:

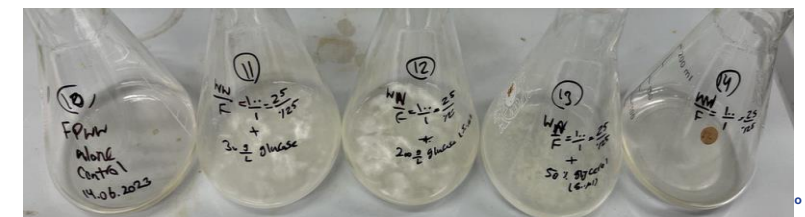
- Fungi thrive on carbon compounds.
- Wastewater environment benefits from fungal activities.

### Practical Implications:

- Enhanced wastewater treatment potential.
- Insights for optimizing fungal-wastewater interactions.

**Result: Adding glucose as a carbon source could enhance the synergic between wastewater and fungus.**

Name	TN	TP
<b>Control (FPWW)</b>	<b>52.6</b>	<b>19.1</b>
W/F= 100/1 (spore)	58.7	18.5
<b>W/F= 100/1 (spore) + glucose</b>	<b>12.5</b>	<b>1.26</b>
W/F= 100/1 (spore) + glycerol	12.8	5.78





## Conclusions:

### Effective and Sustainable Solutions:

- Bio-based technologies offer effective microalgae harvesting methods.
- Harness natural processes for separation, reducing energy consumption.
- Promote sustainability by minimizing chemical and energy inputs.

### Diverse Applicability and Scalability:

- Bio-based methods adaptable to various microalgae species and environments.
- Potential for scalable implementation in both small and large operations.
- Address challenges of traditional methods, making harvesting feasible.

### Pathway to Green Innovation:

- Bio-based approaches align with green and eco-friendly principles.
- Reflect growing interest in sustainable practices in various industries.
- Open doors for innovation and collaboration in biotechnology and beyond.

### Acknowledgments:

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**Thank you for your attention!**

